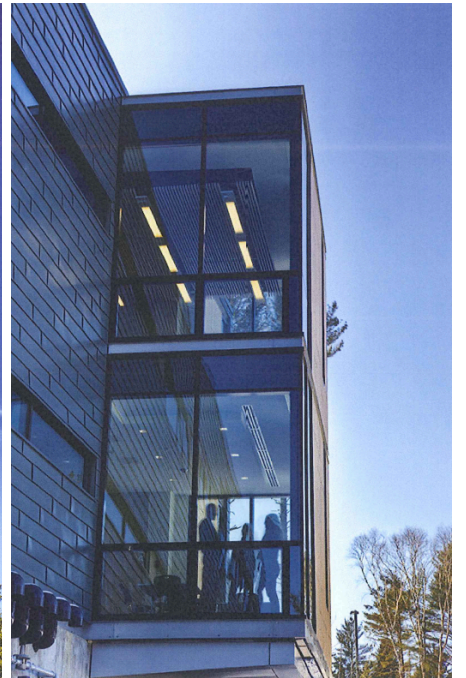
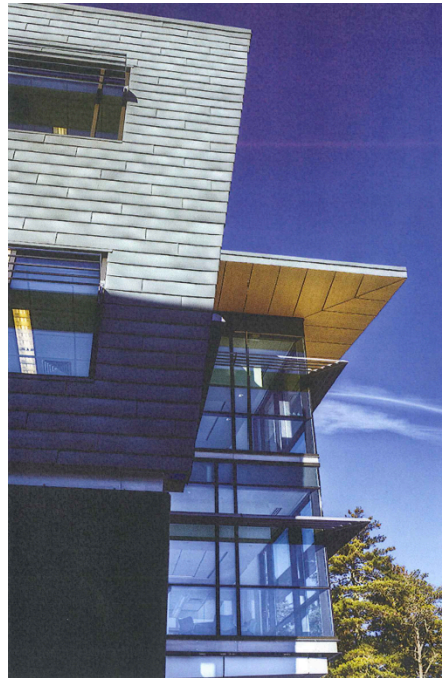
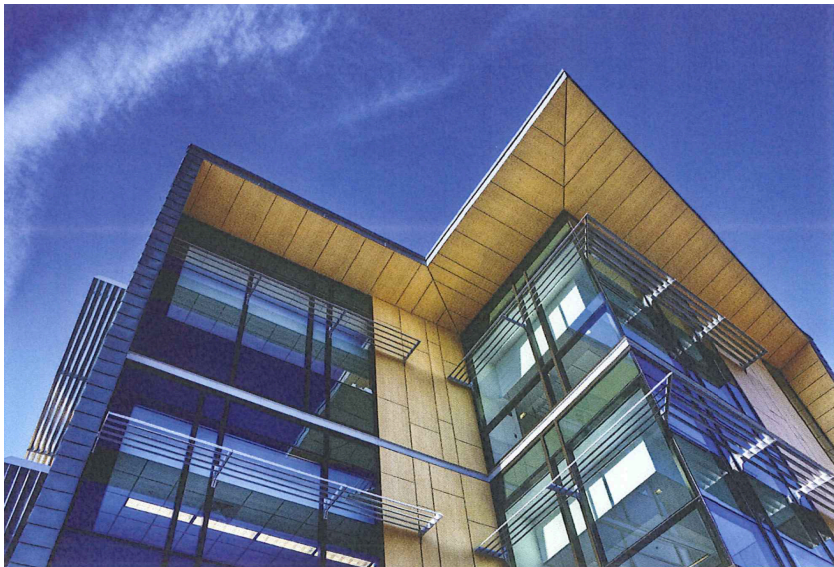
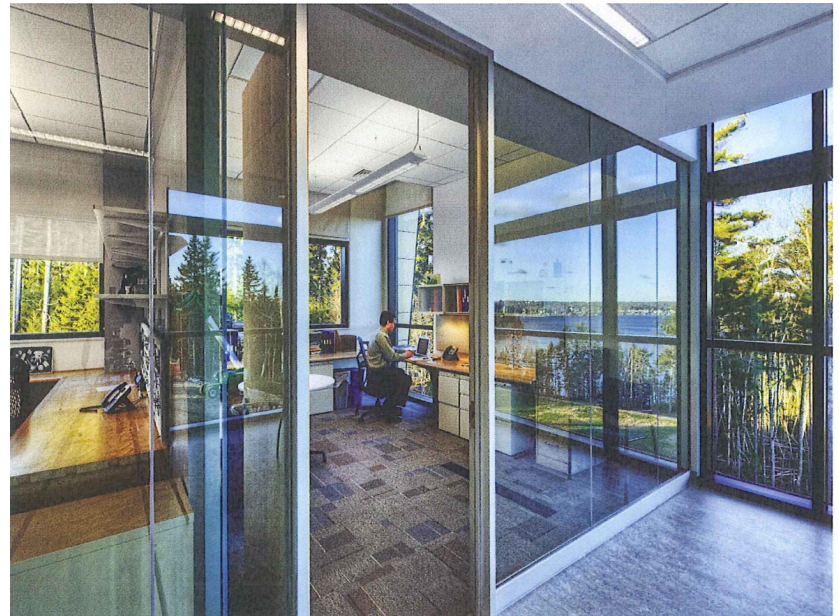
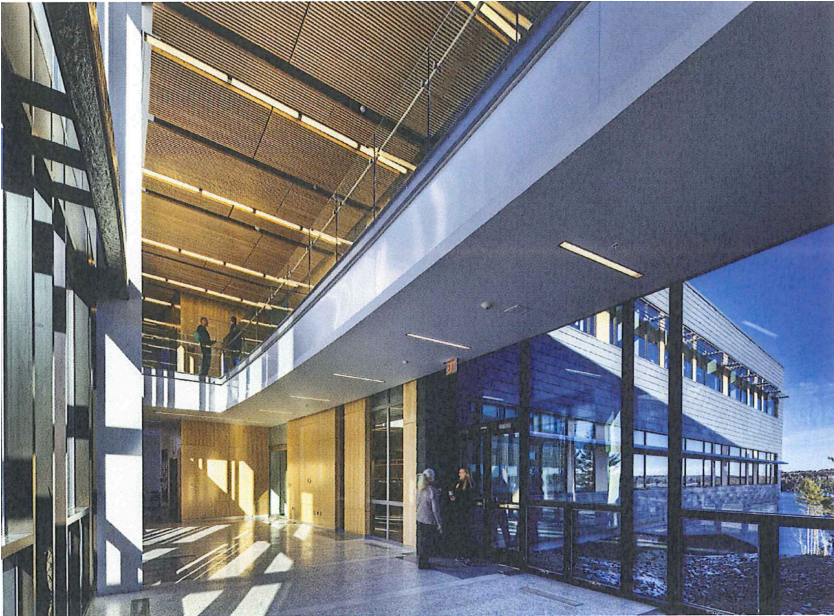




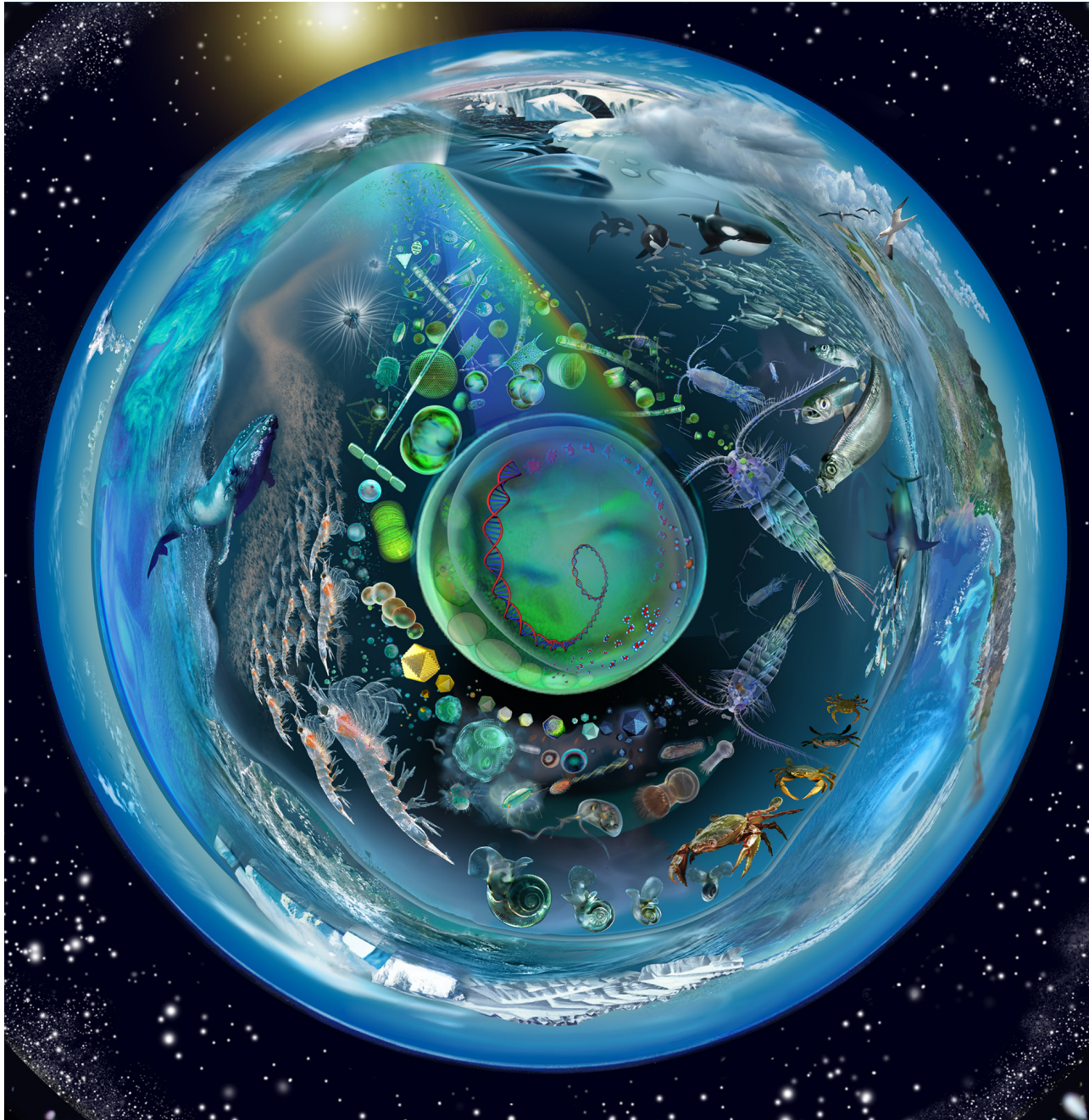
An Introduction to Bigelow Laboratory for Ocean Sciences: Graham Shimmield



Dec 2012

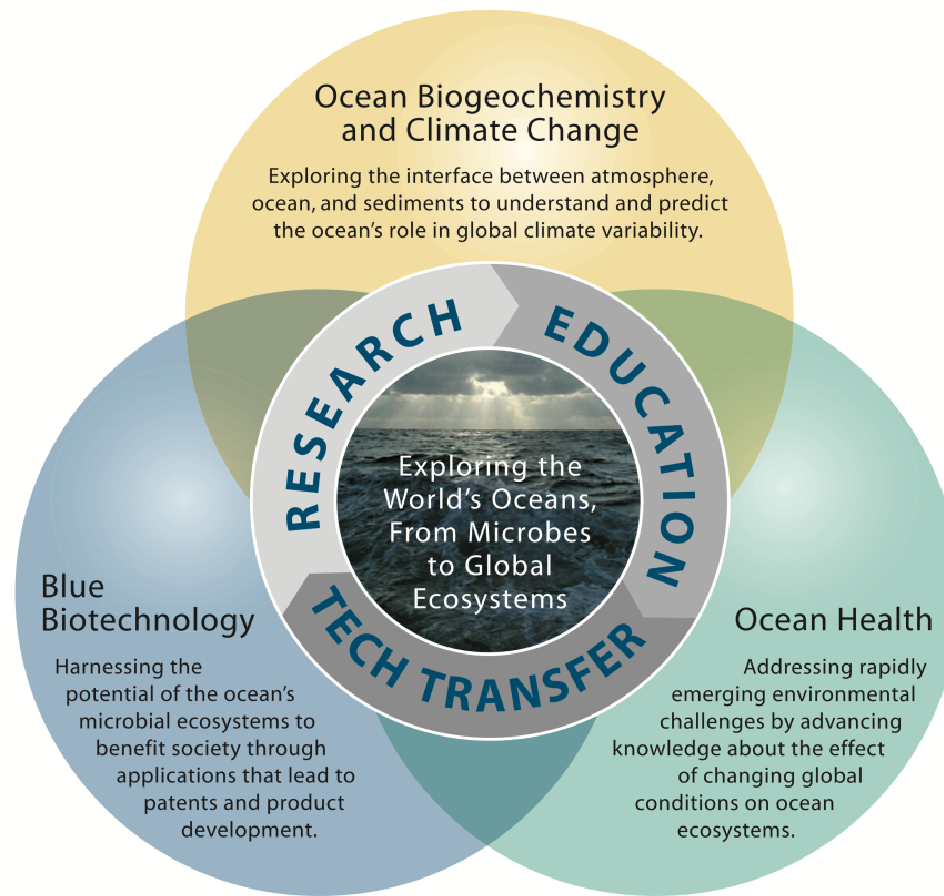






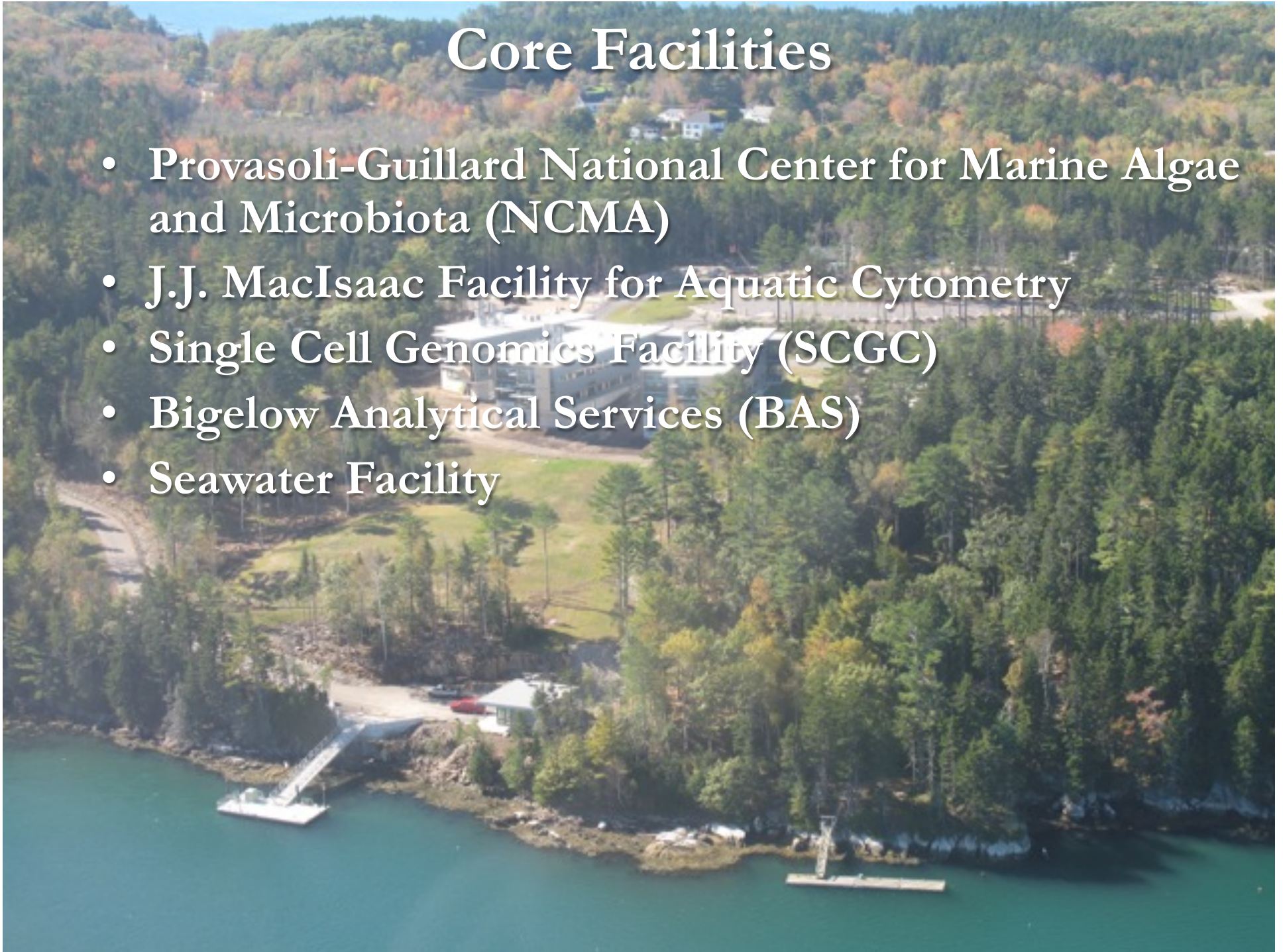
© Bigelow Laboratory for Ocean Sciences; Artist: Glynn Gorick

Core Centers of Discovery: addressing ocean issues in the 21st century



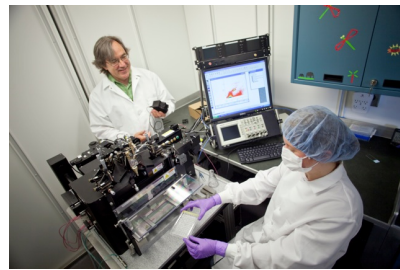
Core Facilities

- Provasoli-Guillard National Center for Marine Algae and Microbiota (NCMA)
- J.J. MacIsaac Facility for Aquatic Cytometry
- Single Cell Genomics Facility (SCGC)
- Bigelow Analytical Services (BAS)
- Seawater Facility



Core Service Centers:

- National Center for Marine Algae and Microbiota
- Single Cell Genomics Center
- Aquatic Flow Cytometry Center
- Bigelow Analytical Services
- Seawater Mesocosm and Large Scale Algal Culture Facility





PROVASOLI-GUILLARD
NCMA
 National Center for Marine Algae and Microbiota

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[Aquaculture Express Strains](#)

[Algal Medium Recipes](#)

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Catalog

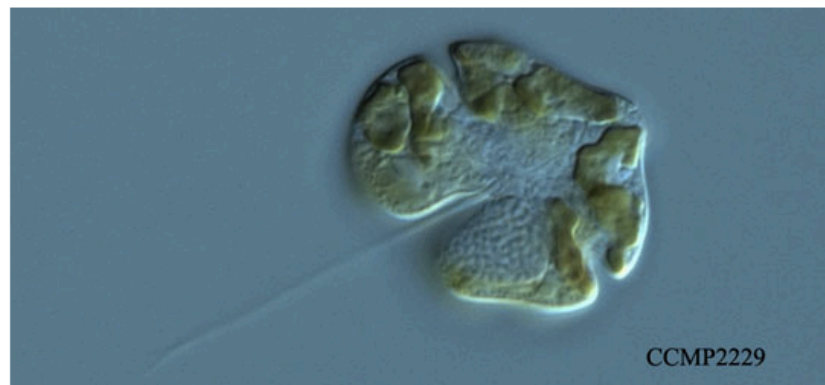
[Medium Kits \(12\)](#)

[Prepared Medium \(17\)](#)

[Seawater \(5\)](#)

[Algal Strains \(2725\)](#)

[Bacterial Strains \(74\)](#)



CCMP2229

NCMA (Formerly the CCMP) - A National Center

We have moved! Please note our new address:

60 Bigelow Drive, PO BOX 380, East Boothbay, Maine 04544

Phone: (001) 202 747 3255 x202

Fax : (001) 202 747 3258

The NCMA is the national marine phytoplankton collection, now incorporating bacteria and viruses, and it is an integral part of Bigelow Laboratory for Ocean Sciences. The NCMA maintains over 2700 strains from around the world, most are marine phytoplankton but we also have benthic, macrophytic, freshwater and heterotrophic organisms.

You can search our [online catalog](#) for strains using taxonomic, geographic and other parameters. Strain records have (when available):

- [collection and isolation information](#)

Latest News & Announcements



Vote for Willie Wilson to the Algal Biomass Organization (ABO) Board of Directors

"In my role as Director of the NCMA I have an excellent overview of the algal industry through my interactions with a wide range of industry and academic clients, all interested in the basics of growing algae for their respective ventures"

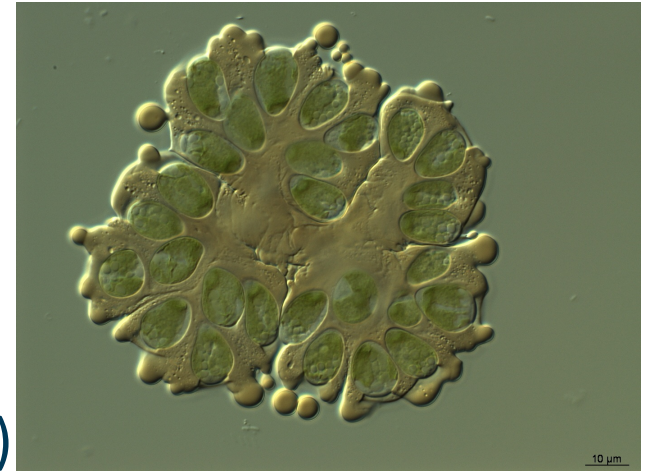
NCMA: A Snapshot

- The world's largest and most diverse living archive of marine microalgae.
- 35 years in the algae business.
- Started as a algal seed stock for the aquaculture industry.
- A repository for public and private collections of algae.
- 2,720 strains of marine, brackish, hypersaline and freshwater algae; including cyanobacteria and macroalgae.
- 359 genera and 723 species.
- 6 products available for every strain.
- 5 growth temperatures (polar to tropical).
- On-site & off-site back-up and cryopreservation.
- Expansion to include new Bacteria and Virus collections.
- New web site to enhance customer experience (Oct 2013).



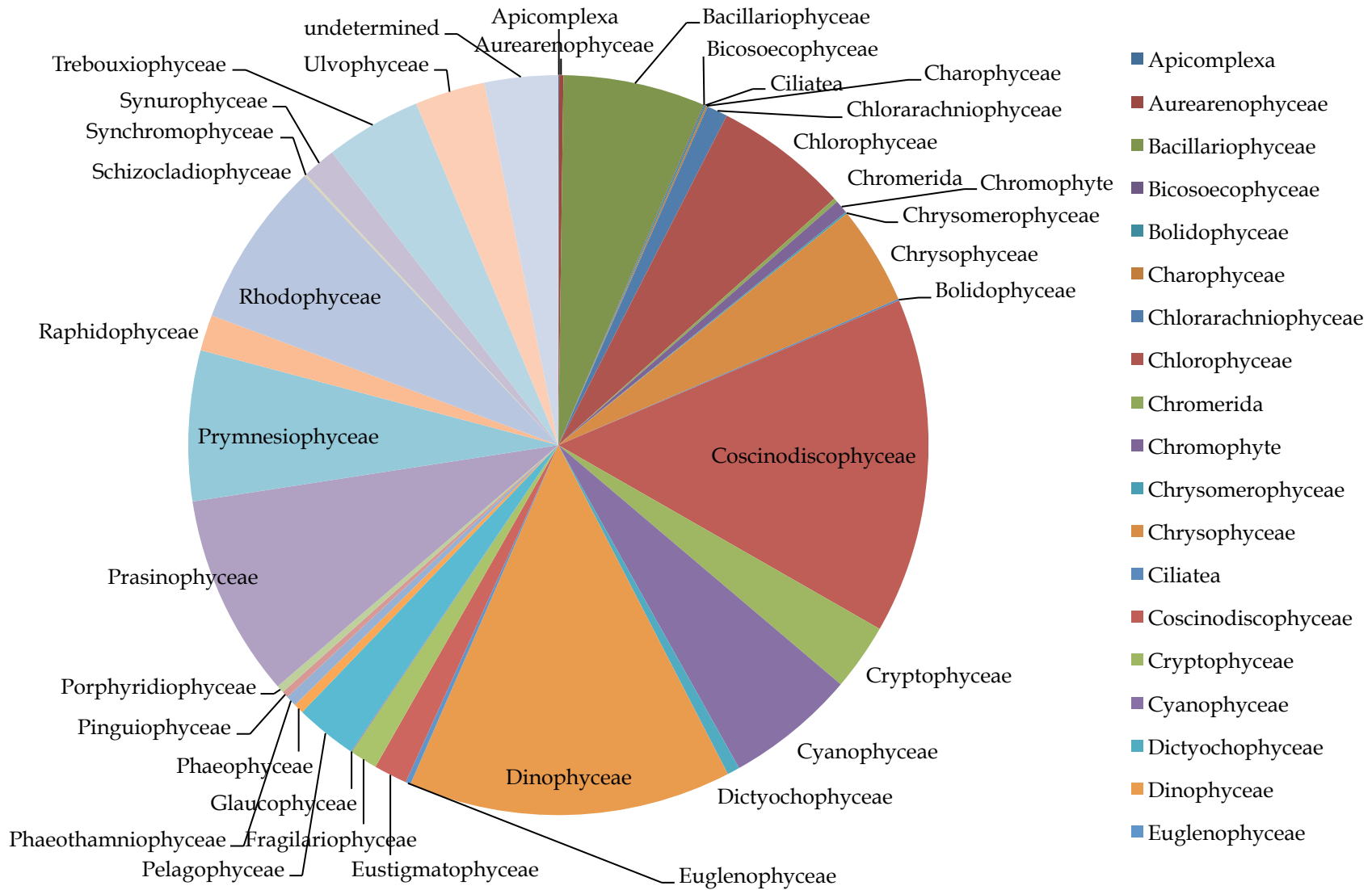
Algal products

- Health food (\$2.5-billion/year)
- Aquaculture (\$700-million/year)
- Animal feed additives (\$300-million/year)
- PUFAs (DHA) (\$1.5-billion/year) (eg Martek)
- Anti-oxidants (eg. Beta-Carotene) (\$400-million/yr)
- Coloring substances (eg. Astaxanthin) (\$160-million/yr)
- Fertilizers/soil conditioners (\$5-billion/yr)



Exploring the World's Oceans, from Microbes to Global Ecosystems

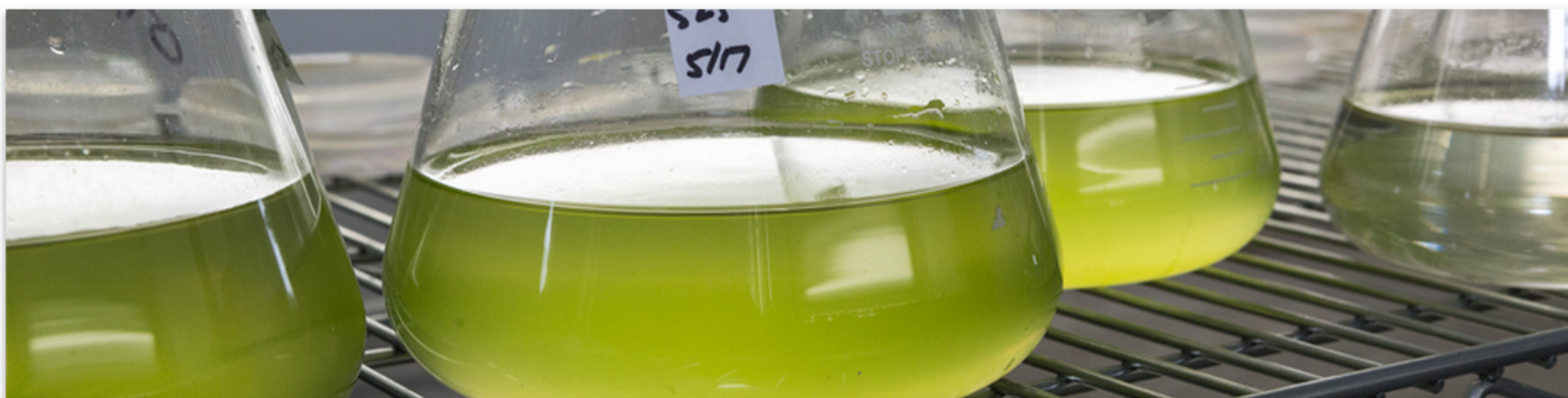
The NCMA holds representatives from 39 Classes of Algae (all the major photosynthetic groups)



Products and Services

- Starter cultures
- Nucleic acids from algae
- Culturing Techniques Courses
- Private collections
- International Depository Authority (patent depository)
- got algae? T-shirts!
- Research services
- Isolations/clean up/taxonomic ID
- Services through our partners (e.g. powdered algae)
- Other service centers at Bigelow
 - Single Cell Genomics
 - Flow Cytometry
 - Analytical Services
 - Seawater Facility



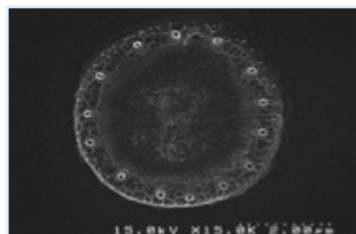


Blue Biotechnology: Harnessing the ocean's potential to benefit society.

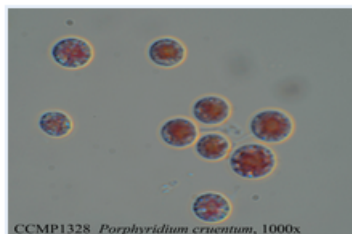


CCMP908

Featured Product/Service:
Aquaculture Express
Strains



Featured Strain:
CCMP1335, *Thalassiosira*
pseudonana (aka 3H)



CCMP1328 *Porphyridium cruentum*, 1000x

Featured Paper: Genome of
the red alga *Porphyridium*
purpureum

Latest News & Announcements

Innovator of the Year Award

Governor Paul LePage presented Bigelow Laboratory for Ocean Sciences with the Maine International Trade Center's (MITC) 2012 Innovator of the Year award

[Read More](#)

7th Annual Algae Biomass Summit

The NCMA is an official sponsor of the 7th annual



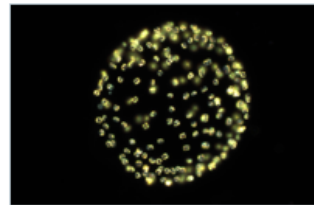
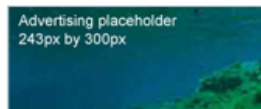
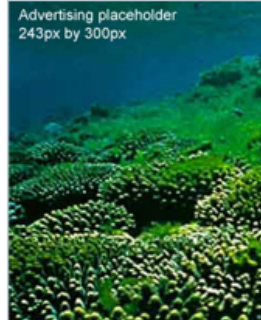
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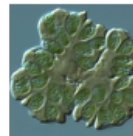
Browse By

Category
[All Algae \(2715\)](#)
[Marine \(898\)](#)
[Freshwater \(308\)](#)
[Hypersaline \(50\)](#)
[Heterotrophic and Mixotrophic \(0\)](#)
[Robust \(81\)](#)
[Aquaculture \(0\)](#)
[Toxic \(303\)](#)
[Cold Water \(224\)](#)
[Warm Water \(51\)](#)
[Temperate \(72\)](#)
[High Lipid \(0\)](#)
[Seaweed \(125\)](#)



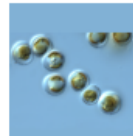
Algae

The NCMA is the largest and most diverse collection of marine algae in the world. With almost half of the cultures in cryopreservation, our curators perpetually culture over 1500 strains. Hundreds of new strains are accessed into the collection each year. The collection is searchable by the Tree of Life, the world map or the following categories.



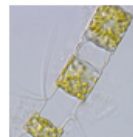
High Lipid

Strains that can be induced to accumulate



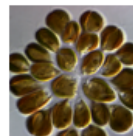
Marine

Strains isolated from the ocean or



Cold Water

Strains that thrive in temperatures less than



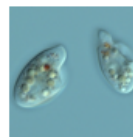
Freshwater

Strains that grow in freshwater media. They



Aquaculture

The NCMA aquaculture express starter



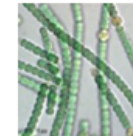
Heterotrophic and Mixotrophic

Heterotrophs require an organic



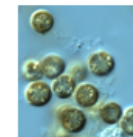
Seaweed

This grouping is based on a generic term



Hypersaline

Strains from an environment with saline



Warm Water

Strains that are grown at temperatures greater than



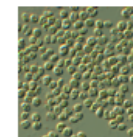
Temperate

Strains that thrive at temperatures between



Toxic

Strains that represent species that are known to

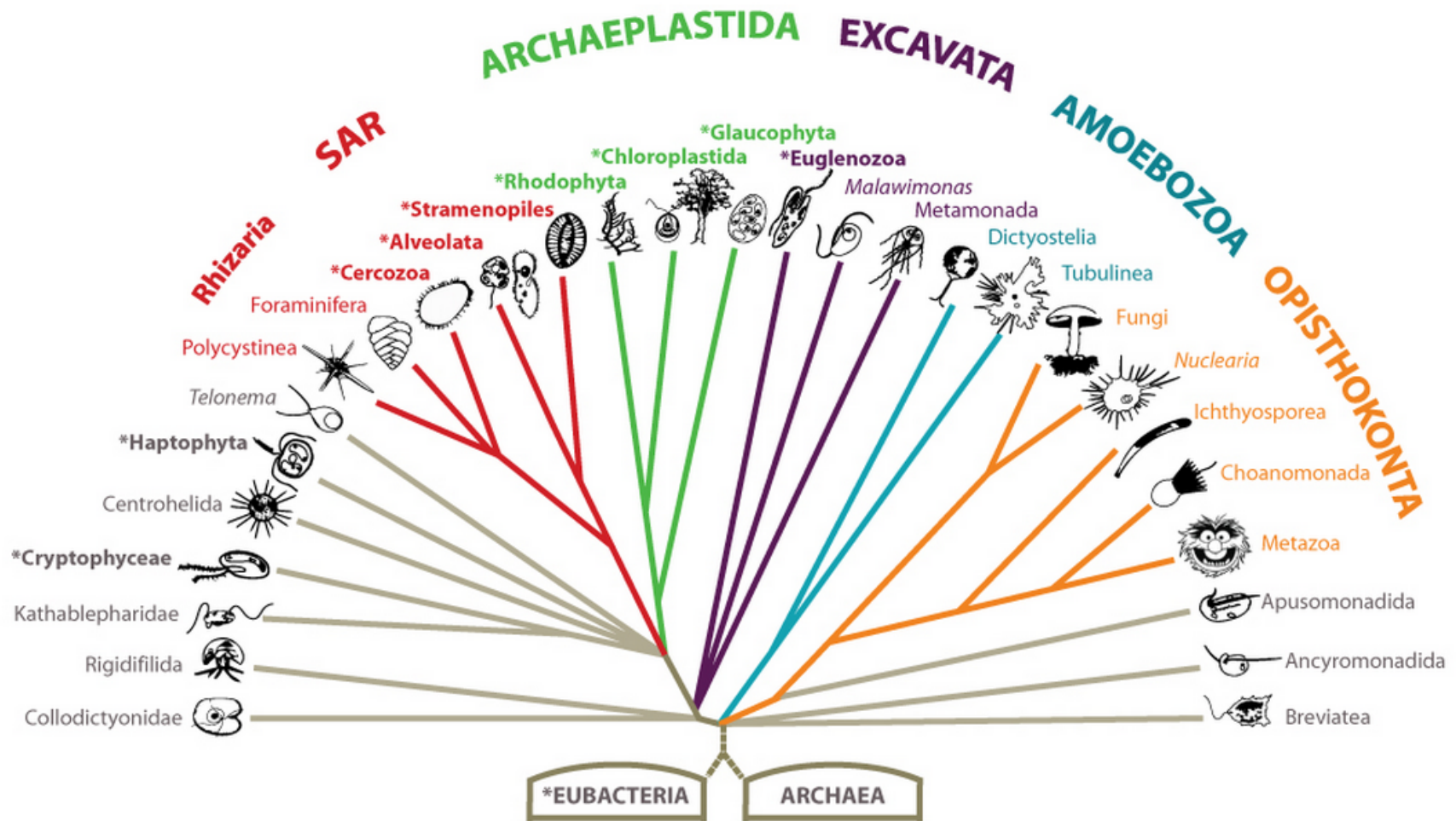


Robust

Strains that grow vigorously, do not die

Interactive Tree of Life

Tree of Life



Google Maps

Geographic Map Locator

Enter Location to narrow down your search:

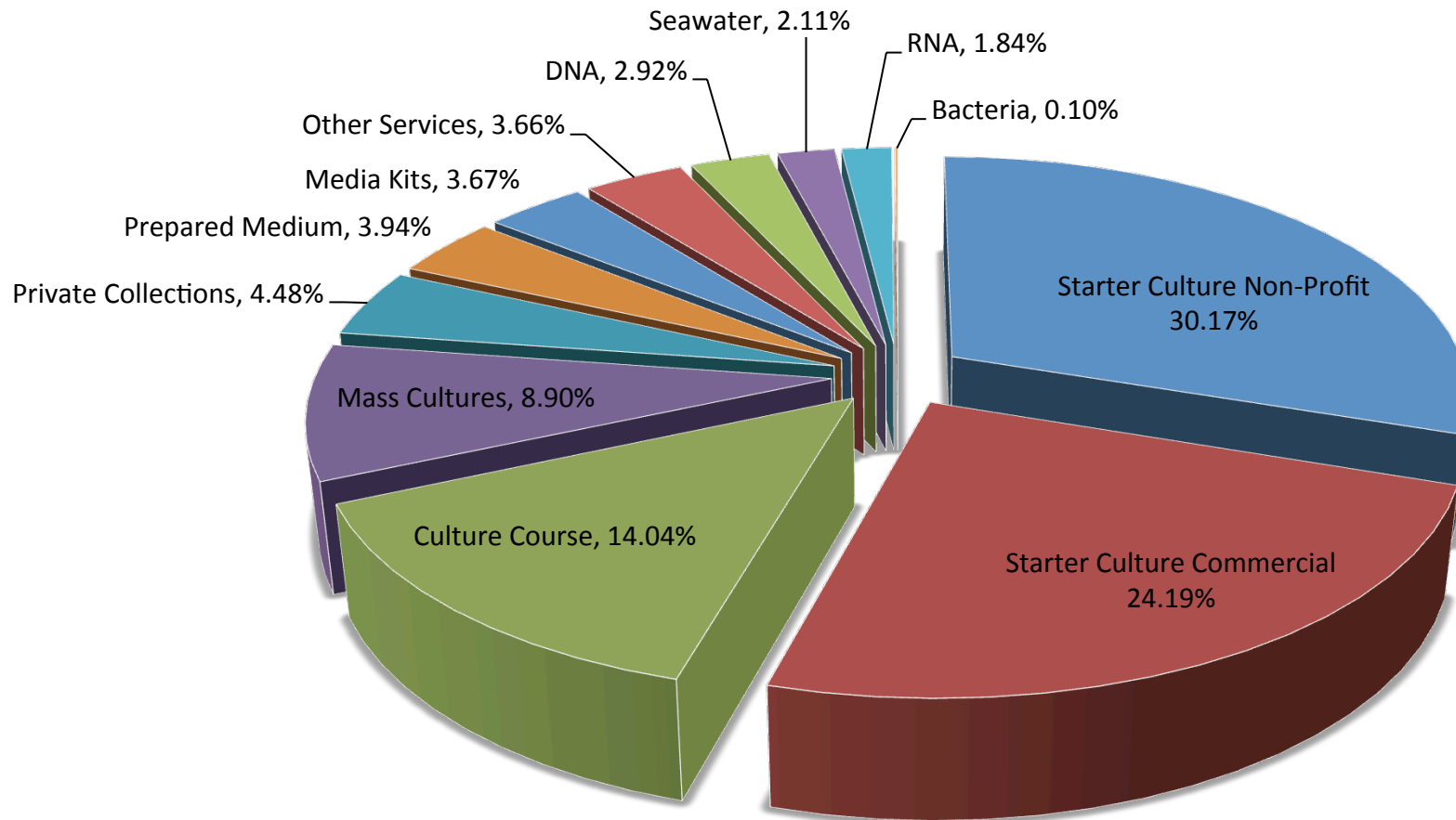
Radius: 200 mi

Filter Map by Tag: [Show all](#) | [Algae](#) [Cyanophyceae](#) [Prasinophyceae](#) [Trebouxiophyceae](#) [Fragilariophyceae](#) [Coscinodiscophyceae](#) [Bacteria](#)

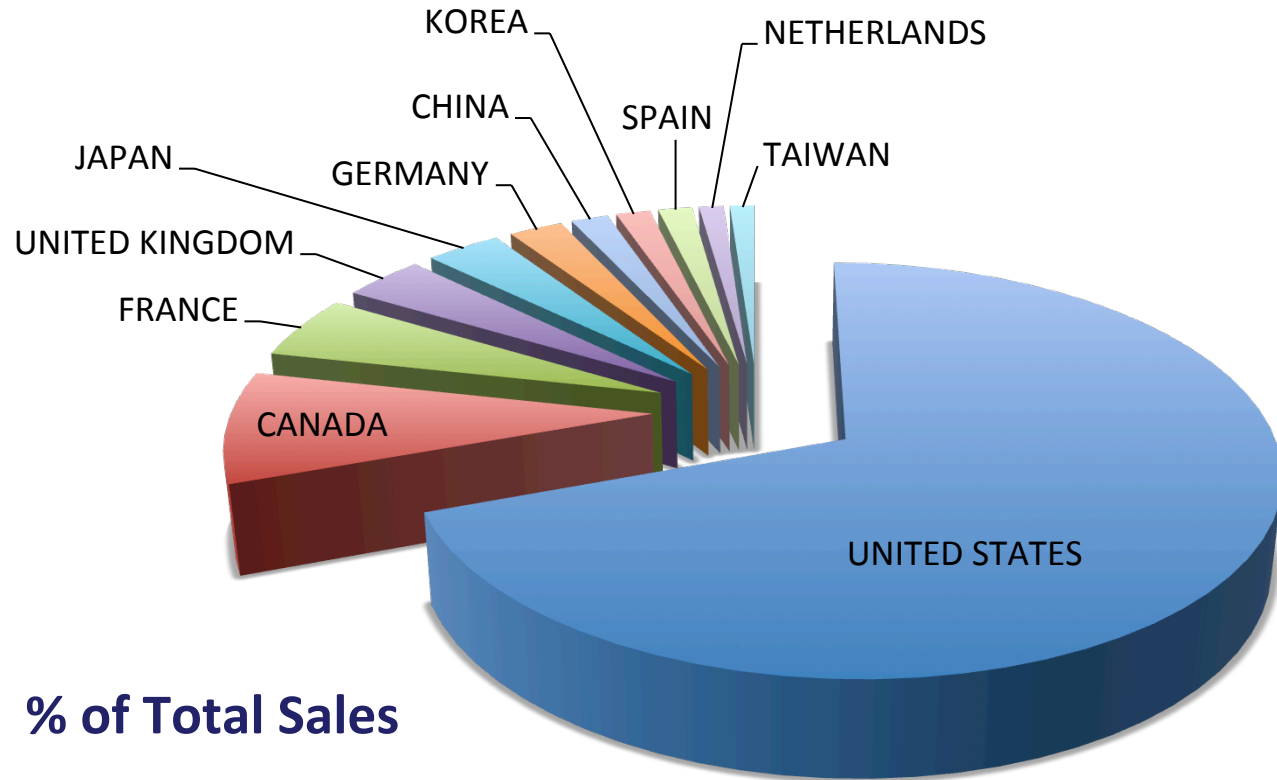
[Reset locations](#)



Percent of Gross Revenue by Product FY2009-2012



Percent of Gross Sales by Country (FY2009-2012)



Algae Attributes

Algae

← Save + × [Icons] ?

Strain Number CCMP1004

Class Coscinodiscophyceae

Genus Thalassiosira

Species oceanica

Variety

Authority (Hustedt) Hasle et Heimdal

Identified by

Axenic by

Axenic date / /

Deposited date 1/14/1983 Deposited by Brzezinski,M

Isolation date / / Isolated by Brzezinski,M

Rack location GD2

Common name

Name synonyms

Strain synonyms 47, NEPCC618

Authentic No Genome?

Strain URL <http://ncma-dev.bielow.org/ccmp1004>

Microscope <http://starcentral.mbl.edu/microscope/portal.php?page=taxonfactsheet&type=>

Valid GenBank record? GenBank URL <http://www.ncbi.nlm.nih.gov/sites/entrez?db=nucleotide&>

Deposited for patent?

Private collection?

Private strain # Customer #

Collected by Brzezinski,M

Collected date 9/26/1981

Collection latitude 40.3923N Collection longitude 63.9570W

Collection site Gulf Stream Warm Core Ring B1D

Ocean North Atlantic Sea

Hydro ecology 33meters,temp 21°C ,downwelling light intensity 8percent I.O.

Nearest continent Open ocean Country

Other

Tree of Life Lvl 1 Sar

Tree of Life Lvl 2 Stramenopiles

Comment He kept in L/20 at 20°C;17Dec09-ID confirmed by Moriz,M and Kaczmarek in "Barcoding of Diatoms:Nuclear Encoded ITS Revised"; Protist, 13Aug2009

Primary growth medium

Best temp range 22-26°C Maintained at NCMA 24 degrees C

Min temp 22 Max temp 26

Min length 6 Max length 20

Min width 5 Max width 10

Min salinity 0.00 Max salinity 0.00

Morphological data

Toxic? Heterotrophic? Bioluminescent?

Aquaculture strain? Marine? Freshwater?

Hyper-saline? Robust? High lipid?

Seaweed? Cold Water? Warm Water?

Temperate?

Growing instructions

Cryopreserved? Failed Last Cryo Date 3/13/2003 DNA barcode?

Cryo Info CCMP1004 was cryopreserved on Mar 13 2003 using 12% DMSO as a cryoprotectant. The time required to regrow this culture, prior shipping, is approximately 67 days. If interested, please contact the CCMP for the cryopreservation methods (freezing and/or thawing protocols). Note that aquaculture strains are always maintained as actively growing cultures, even if also cryogenically stored. Therefore, aquaculture strains (see aquaculture

Temp out date / /

Temp out? Temp out by

Comments (temp out)

Keywords

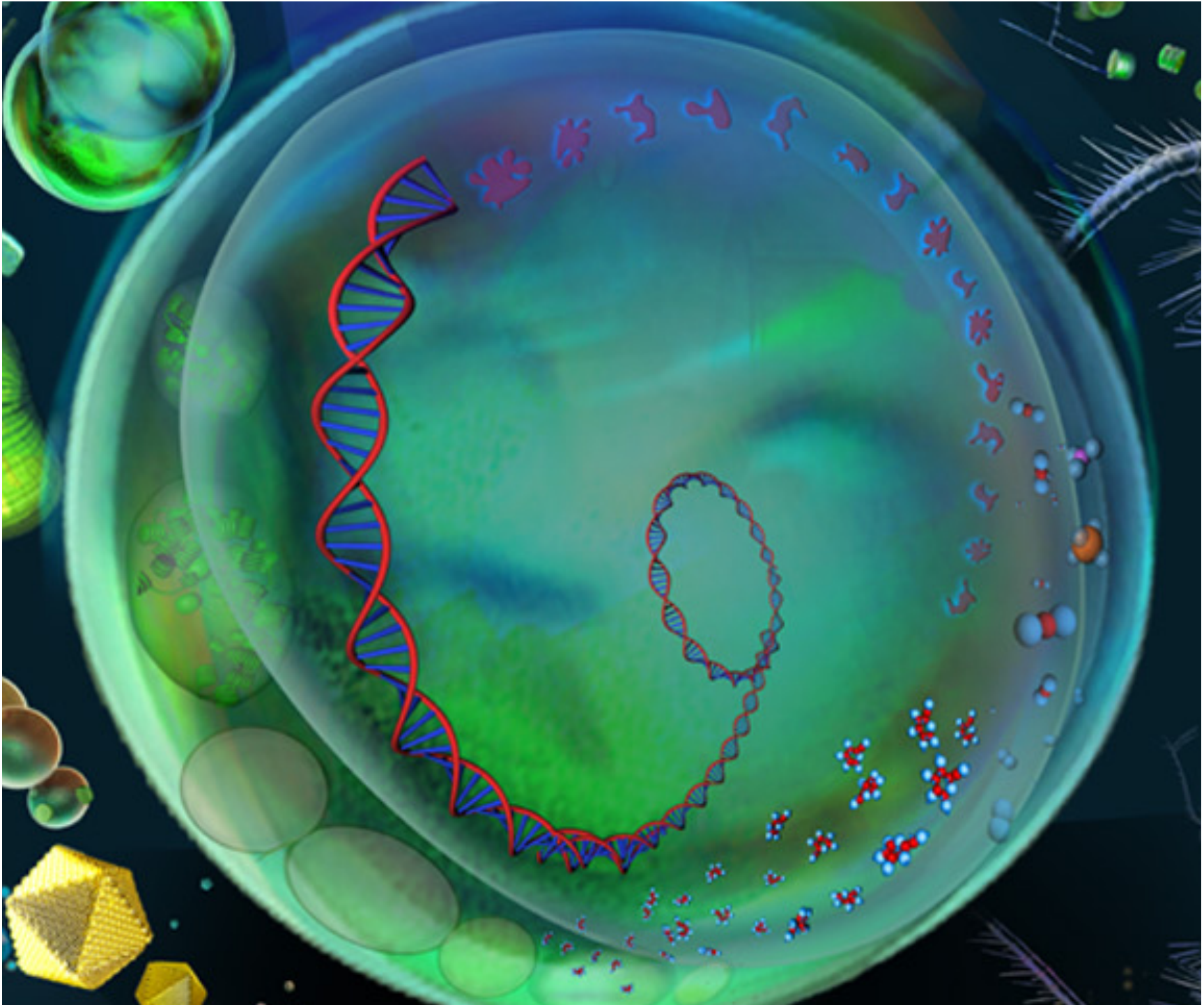
Notes

Deleted? Deleted date / /

Deleted by

Deleted comment

Lot Culture medium Axenic Temp out history Item files

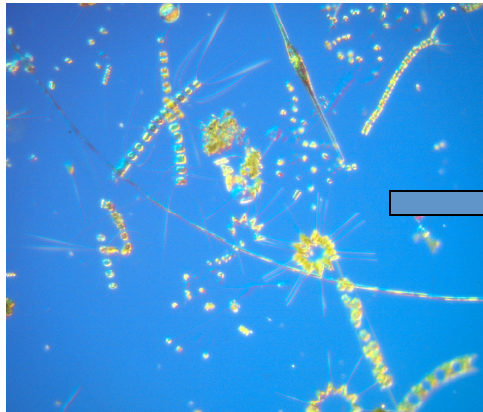


Microbial diversity:

Each mL of ocean water contains $>10^5$ cells
or ~ 1 terabyte of microbial genetic information!



The metagenomics approach



Extract all the
DNA from
microbial
community

Determine what the genes ARE:
(**Sequence-based metagenomics**)

Identify genes and metabolic pathways

“Who, what and why?”

Requires screening against reference libraries

Determine what the genes DO:
(**Function-based metagenomics**)

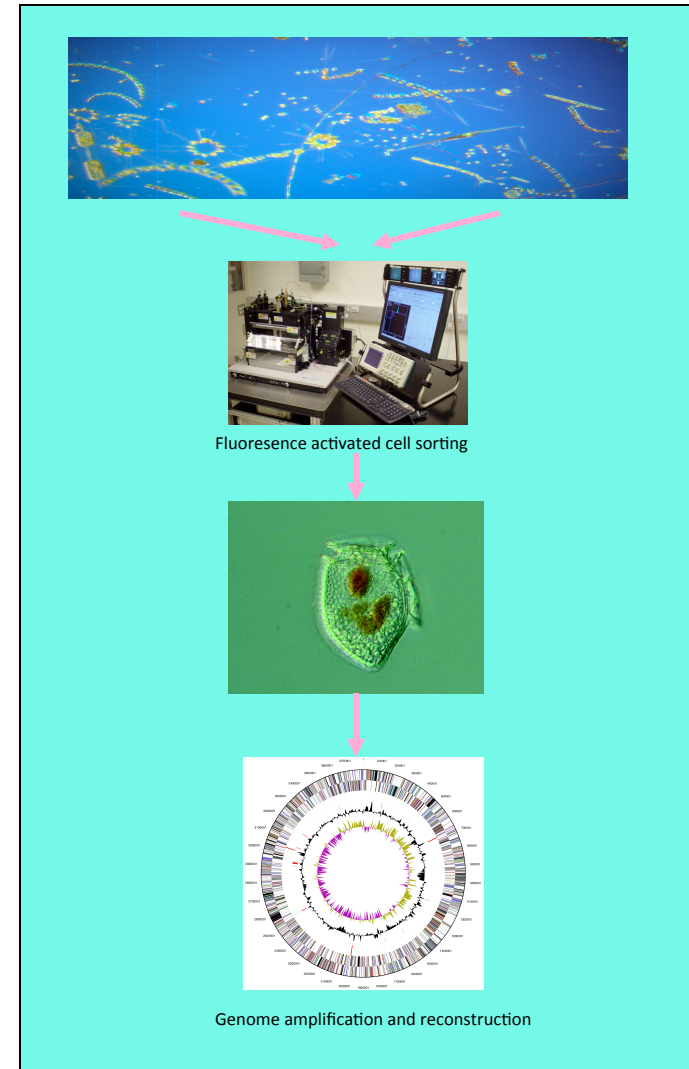
Screen to identify functions of interest, such
as vitamin or antibiotic production

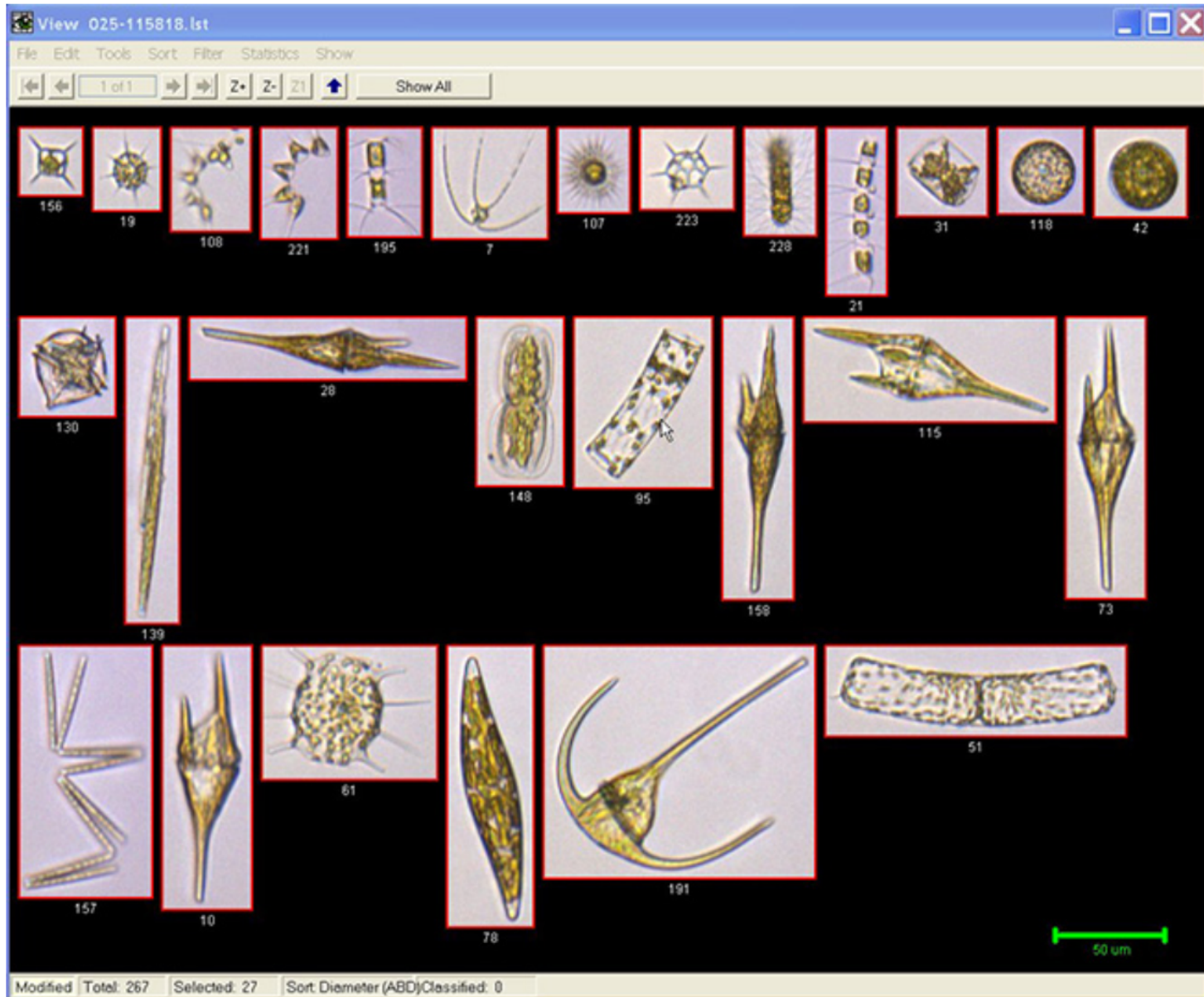
Find the genes that code for functions of
interest

Usually, unambiguous assignment of gene
function

Single cell genomics

- Pioneering research led by Bigelow investigators (Stepanauskas, Sieracki et al)
- Applying the power of **single cell sorting** and **genomics** together.
- Addresses the limitations of shotgun sequencing and uses vital information on the physiology, environmental factors and evolution to **address microbial diversity in the ocean.**





1

Nice Harbor, 10X

Nice, France

Particle Sorting with Flow Cytometry





BIGELOW LABORATORY SINGLE CELL GENOMICS CENTER

Reading nature's genetic tales, one cell at a time



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News & Publications

Education

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High-throughput single cell separation, DNA amplification, sequencing and bioinformatics

Announcements & Events:

UPCOMING CELL SORTING SESSIONS:

- November 19th - 22nd, 2013
- January 21st - 24th, 2014

Our new website is unveiled, please [tell us](#) what you think!

Recent instrumentation awards by [Illumina](#) and [NSF](#) will boost SCGC's genomic sequencing capacity

Bigelow Laboratory receives [State of Maine Innovator of the Year](#) and top [AGC Build New England](#) awards

Single cell genomics technology and applications are poised for [rapid growth](#)

Recent SCGC-enabled discoveries:



[Biology's Dark Matter Illuminated](#)

A collaborative study led by the U.S. Department of Energy Joint Genome Institute sheds light on metabolic features, evolutionary histories and global distribution of 28 major branches of Bacteria and Archaea that have been largely unknown to date, due to their evasion from laboratory cultivation.

Original publication: [Rinke et al. 2013; Nature 499:431-437](#)



[Resourceful Microbes Reign in World's Oceans](#)

In the first large-scale single cell genomics project, a research team led by SCGC scientists discovered that marine microbes are adapted to very narrow and specialized niches in their environment. This may explain why so few of these microbes—usually less than 1%—can be grown for study in the laboratory.

Original publication: [Swan et al. 2013; PNAS 110:11463-11468](#)

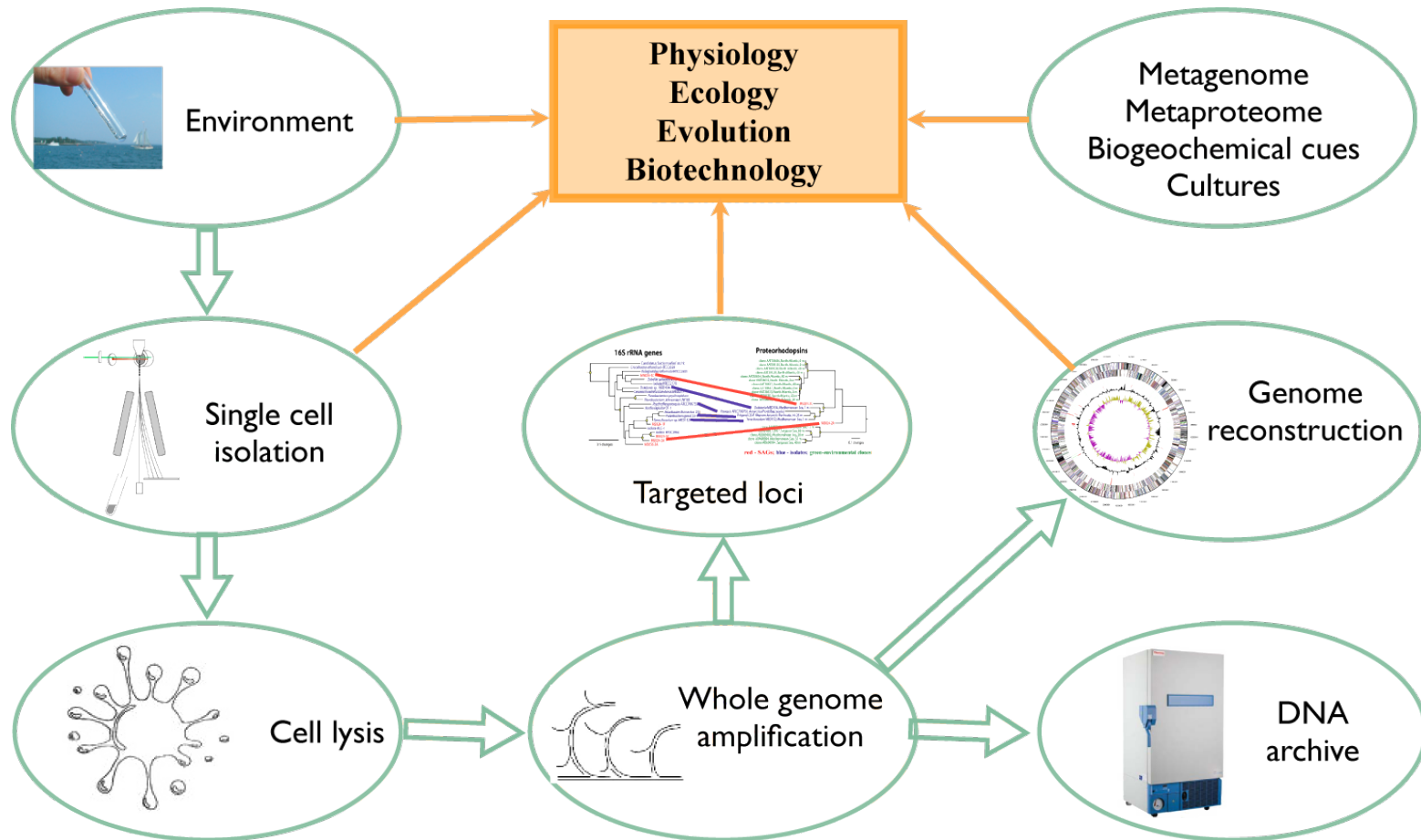


[Intraterrestrial Lifestyles of Marine Archaea Revealed](#)

A study led by scientists from the Aarhus University and the University of Tennessee reveals metabolic features of two lineages of Archaea that are ubiquitous in marine sediments worldwide, suggesting that they may play a key role in the global degradation of buried protein.

Original publication: [Lloyd et al. 2013; Nature 496:215-218](#)

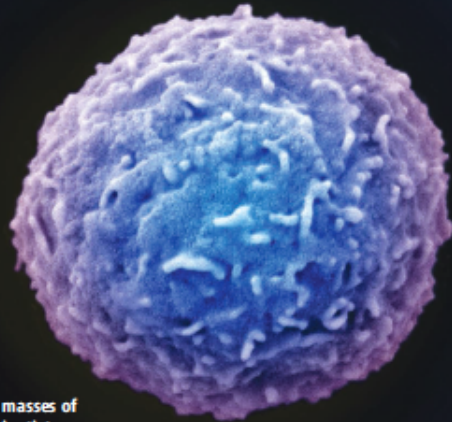
Single Cell Genomics Pipeline



CELL BIOLOGY

The Power Of One

Rather than probe masses of cells all at once, scientists are now applying new research techniques to individual cells



- 1) Measure out a heaping portion of cells.
- 2) Grind until thoroughly mixed.
- 3) Analyze.

Cellular studies still pretty much stick to this traditional recipe, whether the goal is probing bacterial metabolism, following differentiation of stem cells, or tabulating gene activity in tumors. But mashing up a multitude of cells—one common method of studying gene expression typically requires more than 10,000—obliterates key differences between cells, researchers have come to realize. “If you take an average of a large number of cells, you get an average answer,” says analytical chemist Renato Zenobi of the Swiss Federal Institute of Technology in Zurich.

That’s why more and more scientists are opting for the alternative approach of taking the measure of individual cells. Although

much of this work is in its early stages, “there is an increasingly diverse set of examples where single-cell studies have provided qualitative insights that couldn’t be obtained from population-level studies,” says biophysicist Michael Elowitz of the California Institute of Technology in Pasadena.

Scientists have already recorded the most accurate measurements of how much an individual cell weighs and gauged how much oxygen one requires. They’ve flagged specific cancer cells resistant to chemotherapy and developed ways to pinpoint rare, disease-causing bacteria among swarms of harmless microbes. Developmental biologists have tallied gene activity as a fertilized egg starts its course of division and specialization, work that might help clarify the factors that spur a cell in the embryo to become one tissue and its seemingly identical next-door neighbor

◀ **Small sample size.** Researchers are exploring new ways of investigating individual cells such as this human white blood cell.

to become something else. And Elowitz and other researchers have spelled out how individual cells not only cope with but actually benefit from “noise,” random fluctuations in their internal and external conditions.

Of course, scientists have paid attention to single cells ever since the first microscopes were invented. What’s changed is that researchers are now applying to individual cells the powerful techniques, including genome sequencing, mass spectrometry, and gene expression analysis, that formerly required batches of cells. “Real biological tissues are complex, and if you want to dissect that complexity and heterogeneity, you have to have tools to do it at the single-cell level,” says biophysicist Stephen Quake of Stanford University in Palo Alto, California.

Good technique

Single-cell research tools range from old standbys to cutting-edge inventions (see sidebar for a sample of methods). Many of them allow researchers to get into what Quake calls “production mode,” analyzing large numbers of individual cells in parallel or over a short period of time. The technology he calls “absolutely central” to the surge in single-cell research is microfluidics, which uses miniaturized networks of channels, valves, pumps, and chambers to control microscopic quantities of liquid. So-called lab-on-a-chip devices combine microfluidic circuits and can perform several analytical steps.

An example of how microfluidics can elucidate single-cell behavior comes from Quake, his Stanford colleague Markus Covert, and their colleagues. Last July in *Nature*, the

Single-Cell Tech Primer

Microfluidics is the hot technique in the single-cell field (see main text). However, it’s just one of the methods that are enabling researchers to delve into individual cells.

• Gene Expression

Many modern gene-expression studies apply the mingled contents of thousands of cells to devices called microarrays that look like glass slides or microchips. But with microfluidic chips, researchers can make the same measurements on

one cell. Take the gene-expression chips from California-based Fluidigm, a company co-founded by Stephen Quake of Stanford University in Palo Alto, California. The devices isolate samples from individual cells and mix them with the chemicals necessary for quantitative polymerase chain reaction, a technique that determines gene activity by measuring how much messenger RNA a gene makes. The company’s most powerful version allows researchers to simultaneously gauge the activity of

96 genes in 96 individual cells, churning through a batch in about 4 hours.

Another new technology, known as RNA-seq, provides an alternative to microfluidic chips for measuring gene activity in one cell. An offshoot of next-generation genome sequencing, the procedure involves converting a cell’s mRNA molecules back into short stands of DNA, sequencing those DNA fragments, and then matching them up with the gene that originally spawned the mRNAs. Last May in *Cell Stem Cell*, molecular geneticist M. Azim

Surani of the University of Cambridge in the United Kingdom and colleagues reported one of the first studies to apply the RNA-seq technique to single cells, revealing the expression of 385 genes in individual embryonic stem cells. Fans of RNA-seq emphasize that it can measure more genes at once than can microfluidic chips. Meanwhile, chip devotees tout their method’s superior speed.

• Flow Cytometry

A classic technique, flow cytometry sifts and counts cells based

Single Cell Genomics: Dr Ramunas Stepanuskas

Downloaded from www.sciencemag.org on June 28, 2011

CREDIT: ANNE WISTON/AMERICAN RESEARCH LABS/UNLIMITED, INC.

Dark ocean autotrophy

Lead postdoc:
Brandon Swan

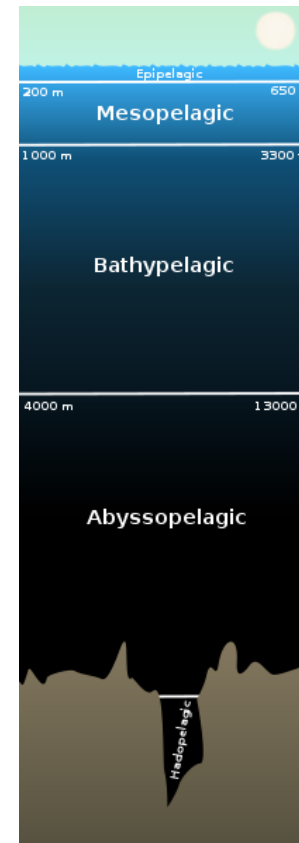
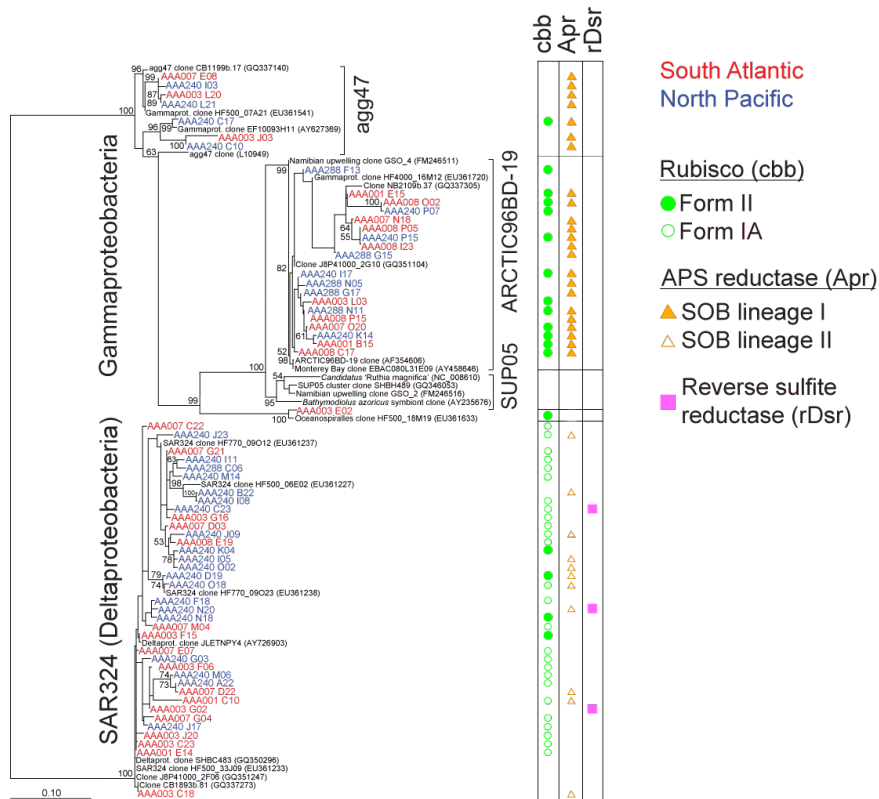


RuBisCO and S oxidation genes detected in SAR324 and γ -Proteobacteria

Consistent in multiple cells from S. Atlantic and N, Pacific

May be significant players in the global carbon cycle

16S rDNA phylogeny and the presence of metabolic genes



Ecology of Picobiliphyta

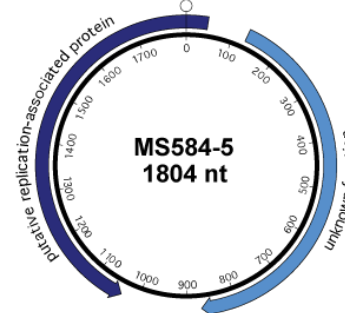
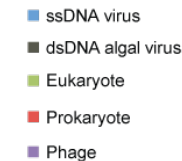
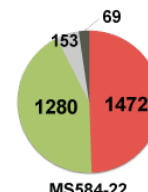
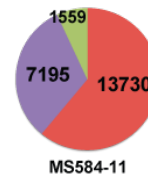
Newly discovered marine protist group

No cultures

Our findings:

- No photosynthesis
- Feed on bacteria and large viruses
- infected by a novel nanovirus

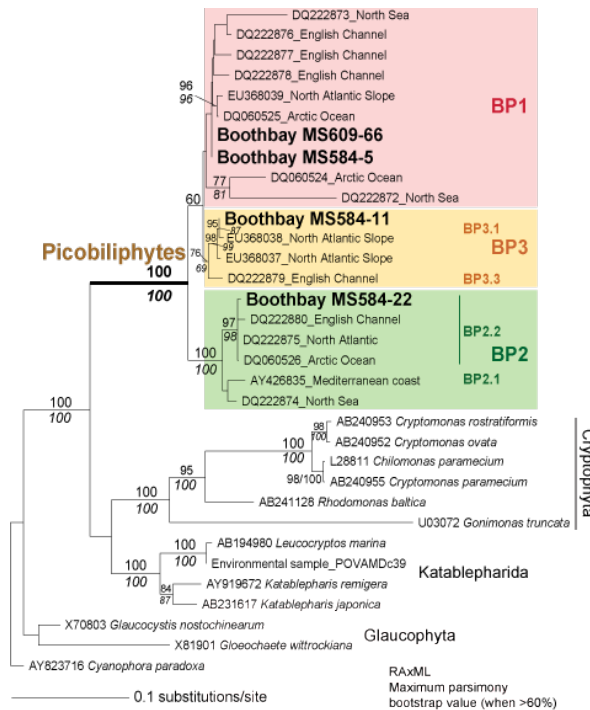
Shotgun read sources



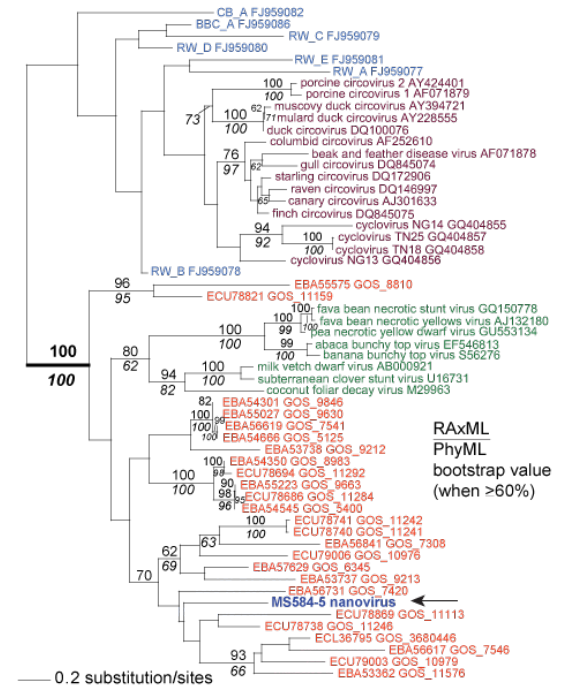
Lead PIs:
HS Yoon (Bigelow)
D Bhattacharya (Rutgers U)



18S rDNA phylogeny



Picobiliphyta-infecting nanovirus



Genomic Encyclopedia of Bacteria and Archaea (GEBA)

led by T Woyke (DOE JGI)

Goals:

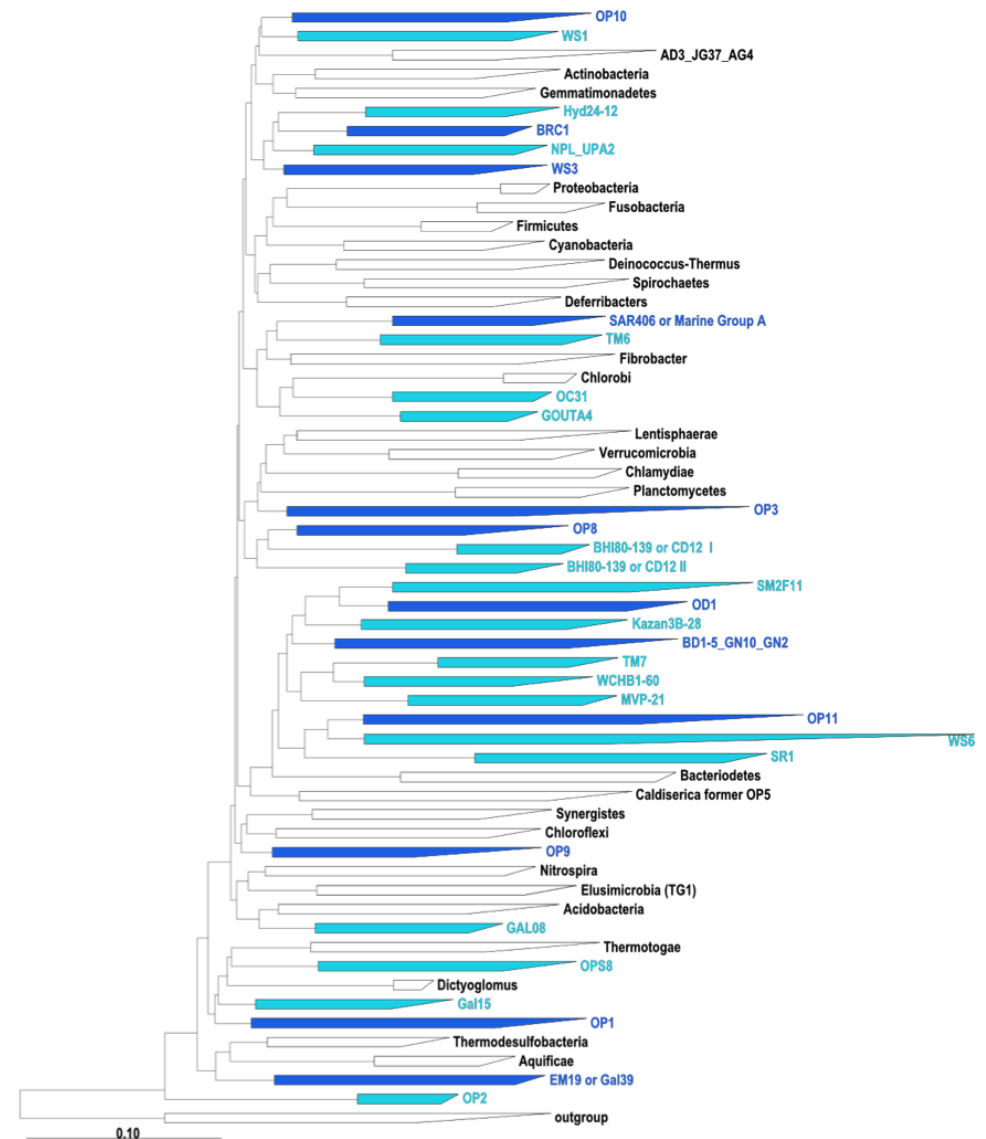
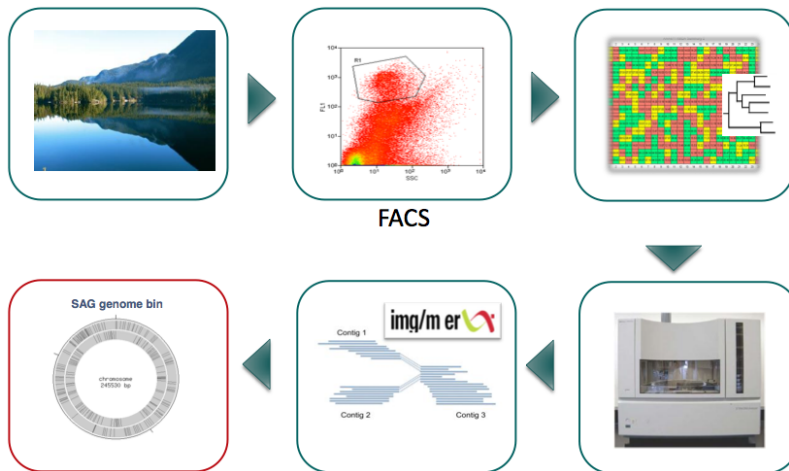
- Representing each major branch of the tree of life with a genome
- Discovering of genes, proteins and pathways
- Resolving early evolution of life

Methods:

- Single cell FACS and MDA (Bigelow SCGC)
- Whole genome sequencing (JGI)

Uncultured groups represented so far:

- ~15 Bacteria phyla
- ~10 Archaea phyla



Prevalent genome streamlining and latitudinal divergence of planktonic bacteria in the surface ocean

Brandon K. Swan^a, Ben Tupper^a, Alexander Szczyrba^b, Federico M. Lauro^c, Manuel Martínez-García^d, José M. González^e, Haiwei Luo^f, Jody J. Wright^g, Zachary C. Landry^h, Niels W. Hansenⁱ, Brian P. Thompson^j, Nicole J. Poulton^k, Patrick Schwientek^l, Silvia G. Acinas^m, Stephen J. Giovannoniⁿ, Mary Ann Moran^o, Steven J. Hallam^p, Ricardo Cavichio^q, Tanja Woyke^r, and Ramunas Stepanauskas^{s,*}

^aSigelow Laboratory for Ocean Sciences, East Boothbay, ME 04544; ^bCenter for Biotechnology, Bielefeld University, 33615 Bielefeld, Germany; ^cSchool of Biotechnology and Biomolecular Sciences, University of New South Wales, Sydney, NSW 2052, Australia; ^dDepartment of Physiology, Genetics and Microbiology, University of Alicante, 03080 Alicante, Spain; ^eDepartment of Microbiology, University of La Laguna, ES-38206 La Laguna, Tenerife, Spain; ^fDepartment of Marine Sciences, University of Georgia, Athens, GA 30602; ^gMicrobiology and Immunology, University of British Columbia, Vancouver, BC, Canada V6T 1Z4; ^hDepartment of Microbiology, Oregon State University, Corvallis, OR 97331; ⁱGraduate Program in Bioinformatics, University of British Columbia, Vancouver, BC, Canada V6T 1Z4; ^jUS Department of Energy Joint Genome Institute, Walnut Creek, CA 94598; ^kDepartment of Marine Biology and Oceanography, Institute of Marine Science, Consejo Superior de Investigaciones Científicas, ES-08003 Barcelona, Spain

Edited by W. Ford Doolittle, Dalhousie University, Halifax, Canada, and approved May 28, 2013 (received for review March 7, 2013)

Planktonic bacteria dominate surface ocean biomass and influence global biogeochemical processes, but remain poorly characterized owing to difficulties in cultivation. Using large-scale single cell genomics, we obtained insight into the genome content and biogeography of many bacterial lineages inhabiting the surface ocean. We found that compared with existing cultures, natural bacterioplankton have smaller genomes, fewer gene duplications, and are depleted in guanine and cytosine, noncoding nucleotides, and genes encoding transcription, signal transduction, and noncytoplasmic proteins. These findings provide strong evidence that genome streamlining and oligotrophy are prevalent features among diverse, free-living bacterioplankton, whereas existing laboratory cultures consist primarily of copiotrophs. The apparent ubiquity of metabolic specialization and mixotrophy, as predicted from single cell genomes, also may contribute to the difficulty in bacterioplankton cultivation. Using metagenome fragment recruitment against single cell genomes, we show that the global distribution of surface ocean bacterioplankton correlates with temperature and latitude and is not limited by dispersal at the time scales required for nucleotide substitution to exceed the current operational definition of bacterial species. Single cell genomes with highly similar small subunit rRNA gene sequences exhibited significant genomic and biogeographic variability, highlighting challenges in the interpretation of individual gene surveys and metagenome assemblies in environmental microbiology. Our study demonstrates the utility of single cell genomics for gaining an improved understanding of the composition and dynamics of natural microbial assemblages.

comparative genomics | marine microbiology | microbial ecology | microbial microevolution | operational taxonomic unit

Planktonic bacteria dominate surface ocean biomass and have a major impact on the global cycling of carbon, nitrogen, and other elements (1). Among the available pure cultures of marine bacterioplankton, only a limited number represent bacterioplankton that are abundant in the ocean, such as the cyanobacteria *Prochlorococcus* and *Synechococcus* and the Alphaproteobacteria *Pelagibacter* (collectively termed PSP cultures). This limits the scope of studies of the microbial metabolic processes and evolutionary changes that impact marine ecosystems and their geochemical cycles (2–6). Unusual nutritional requirements resulting from genome reduction may contribute to cultivation difficulties, as suggested by studies of the chemotetrotroph *Pelagibacter* (7, 8) and the methylotroph OM43 (9).

Although prevailing culture-independent tools, including microbial community shotgun sequencing, targeted gene surveys, and fluorescence in situ hybridization, have revealed the extent and significance of microbial diversity, they have not been able to provide the genome context information required for accurate

metabolic reconstruction spanning organismal, population, and community levels of organization (10). As a result, the genomic repertoire, natural histories, and geographic distribution of even the most abundant taxonomic groups of marine bacterioplankton remain largely unknown (1, 11). Microbial studies in other environments, such as the human body and soils, face similar challenges (10). The recent development of robust protocols for single cell genomics provides a versatile, cultivation-independent approach for assessing natural microbial diversity with corresponding genome context information (12).

To determine whether genome streamlining is a prevalent feature among free-living marine bacterioplankton, and to analyze global patterns of surface ocean bacterioplankton distribution, we obtained draft genomes of 56 single amplified genomes (SAGs) (5, 13–15) and compared them with existing bacterioplankton cultures and metagenomes. The sequenced SAGs represent many ubiquitous surface ocean bacterial lineages, including Marine Group A, *Verrucomicrobia*, *Actinobacteria*, *Bacteroidetes*, and *Proteobacteria* lineages SAR86, ARCTIC96BD-19, SAR92, SAR116, and Roseobacter (*SI Appendix*, Fig. S1). The majority of these groups have few or no cultured representatives. Members of the PSP group were excluded from SAG selection, because their genome streamlining and environmental abundance have been demonstrated previously (1, 2, 4, 11). Samples for SAG generation were collected from the Gulf of Maine, the Mediterranean Sea, and the subtropical gyres of the North Pacific and South Atlantic Oceans (*SI Appendix*, Table S1). On average, 55% (range, 0.3–97.8%) of the genome was recovered from each analyzed cell (*SI Appendix*, Table S2). A subset of 41 SAGs, each >0.75 Mbp in size and with >30% estimated genome recovery, was used for our

Author contributions: B.K.S., B.T., A.S., and R.S. designed research; B.K.S., B.T., A.S., F.M.L., M.M.-G., L.M.G., H.L., J.L.W., Z.C.L., N.W.H., B.P.T., N.J.P., S.G.A., T.W., and R.S. performed research; B.K.S., B.T., A.S., T.W., and R.S. contributed new reagents/analytic tools; B.K.S., B.T., A.S., F.M.L., M.M.-G., L.M.G., H.L., J.L.W., Z.C.L., N.W.H., P.S., S.G.A., S.J.G., M.A.M., S.J.H., R.C., T.W., and R.S. analyzed data; B.K.S. and R.S. wrote the paper.

The authors declare no conflict of interest.

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Data deposition: Whole-genome sequence data for single amplified genomes used for our analyses are available in the Joint Genome Institute's Integrated Microbial Genome database, <http://img.jgi.doe.gov/Woq3-bin/Woq3-bin> (accession nos. 643386079, 643386118, 2223664025–26, 22236664028–29, 22236664032, 22236664034, 22236664035–53, 22236664035–56, 22236664036–37, 22236664038–39, 22236664041, 22236664043, 22236664044, 22236664045, 22236664046, 22236664047, 22236664048, 22236664049, 22236664050, 22236664051–54, 22236664055–56, 22236664059, 22236664061–63, 22236664065–68, 22236664070, 22236664071, 22236664072, 22236664074–76, 22236664080–82, 22236664086–88, 22236664090–92, 22236664094–96, 22236664100–102, 22236664104–106, 22236664108–110, 22236664114–116, 22236664120–122, 22236664124–126, 22236664130–132, 22236664136–138, 22236664142–144, 22236664148–150, 22236664154–156, 22236664160–162, 22236664166–168, 22236664172–174, 22236664178–180, 22236664184–186, 22236664190–192, 22236664196–198, 22236664202–204, 22236664208–210, 22236664214–216, 22236664220–222, 22236664224–226, 22236664230–232, 22236664236–238, 22236664242–244, 22236664248–250, 22236664254–256, 22236664260–262, 22236664266–268, 22236664272–274, 22236664278–280, 22236664284–286, 22236664290–292, 22236664296–298, 22236664302–304, 22236664308–310, 22236664314–316, 22236664320–322, 22236664326–328, 22236664332–334, 22236664338–340, 22236664344–346, 22236664350–352, 22236664356–358, 22236664362–364, 22236664368–370, 22236664374–376, 22236664380–382, 22236664386–388, 22236664392–394, 22236664398–400, 22236664404–406, 22236664410–412, 22236664416–418, 22236664422–424, 22236664428–430, 22236664434–436, 22236664440–442, 22236664446–448, 22236664452–454, 22236664458–460, 22236664464–466, 22236664470–472, 22236664476–478, 22236664482–484, 22236664488–490, 22236664494–496, 22236664500–502, 22236664506–508, 22236664512–514, 22236664518–520, 22236664524–526, 22236664530–532, 22236664536–538, 22236664542–544, 22236664548–550, 22236664554–556, 22236664560–562, 22236664566–568, 22236664572–574, 22236664578–580, 22236664584–586, 22236664590–592, 22236664596–598, 22236664602–604, 22236664608–610, 22236664614–616, 22236664620–622, 22236664626–628, 22236664632–634, 22236664638–640, 22236664644–646, 22236664650–652, 22236664656–658, 22236664662–664, 22236664668–670, 22236664674–676, 22236664680–682, 22236664686–688, 22236664692–694, 22236664698–700, 22236664704–706, 22236664710–712, 22236664716–718, 22236664722–724, 22236664728–730, 22236664734–736, 22236664740–742, 22236664746–748, 22236664752–754, 22236664758–760, 22236664764–766, 22236664770–772, 22236664776–778, 22236664782–784, 22236664788–790, 22236664794–796, 22236664800–802, 22236664806–808, 22236664812–814, 22236664818–820, 22236664824–826, 22236664830–832, 22236664836–838, 22236664842–844, 22236664848–850, 22236664854–856, 22236664860–862, 22236664866–868, 22236664872–874, 22236664878–880, 22236664884–886, 22236664890–892, 22236664896–898, 22236664902–904, 22236664908–910, 22236664914–916, 22236664920–922, 22236664926–928, 22236664932–934, 22236664938–940, 22236664944–946, 22236664950–952, 22236664956–958, 22236664962–964, 22236664968–970, 22236664974–976, 22236664980–982, 22236664986–988, 22236664992–994, 22236664998–1000).

To whom correspondence should be addressed. E-mail: stepanauskas@jgi.doe.gov.

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ARTICLE

Insights into the phylogeny and coding potential of microbial dark matter

Christian Rinke¹, Patrick Schwientek¹, Alexander Szczyrba^{1,2}, Natalia N. Ivanova¹, Iain J. Anderson^{1,3}, Jan-Fang Cheng¹, Aaron Darling^{4,5}, Stephanie Malfatti¹, Brandon K. Swan¹, Esther A. Gies⁶, Jeremy A. Dodsworth¹, Brian P. Hedlund⁷, George Islamis⁸, Stefan M. Slevart⁹, Wen-Iso Liu¹⁰, Jonathan A. Eisen¹¹, Steven J. Hallam⁶, Nikos C. Kyrpides¹², Ramunas Stepanauskas³, Edward M. Rubin¹, Philip Hugenholtz¹¹, & Tanja Woyke¹

Genome sequencing enhances our understanding of the biological world by providing blueprints for the evolutionary and functional diversity that shapes the biosphere. However, microbial genomes that are currently available are of limited phylogenetic breadth, owing to our historical inability to cultivate most microorganisms in the laboratory. We apply single-cell genomics to target and sequence 201 uncultivated archaeal and bacterial cells from nine diverse habitats belonging to 29 major mostly uncharted branches of the tree of life, so-called 'microbial dark matter'. With this additional genomic information, we are able to resolve many intra- and inter-phylum-level relationships and to propose two new superphyla. We uncover unexpected metabolic features that extend our understanding of biology and challenge established boundaries between the three domains of life. These include a novel amino acid use for the *opal* stop codon, an archaeal-type purine synthesis in Bacteria and complete sigma factors in Archaea similar to those in Bacteria. The single-cell genomes also served to phylogenetically anchor up to 20% of metagenomic reads in some habitats, facilitating organism-level interpretation of ecosystem function. This study greatly expands the genomic representation of the tree of life and provides a systematic step towards a better understanding of biological evolution on our planet.

Microorganisms are the most diverse and abundant cellular life forms on Earth, occupying every possible metabolic niche. The large majority of these organisms have not been obtained in pure culture and we have only recently become aware of their presence mainly through cultivation-independent molecular surveys based on conserved marker genes (chiefly small subunit ribosomal RNA; SSU rRNA) or through shotgun sequencing (metagenomes)^{1,2}. As an increasing number of environments are deeply sequenced using next-generation technologies, diversity estimates for Bacteria and Archaea continue to rise, with the number of microbial 'species' predicted to reach well into the millions³. According to SSU rRNA-based phylogeny, these fall into at least 60 major lines of descent (phyla or divisions) within the bacterial and archaeal domains⁴, of which half have no cultivated representatives (so-called 'candidate' phyla). This biased representation is even more fundamentally skewed when considering that more than 88% of all microbial isolates belong to only four bacterial phyla, the Proteobacteria, Firmicutes, Actinobacteria and Bacteroidetes (Supplementary Fig. 1a). Genome sequencing of microbial isolates naturally reflects this cultivation bias (Supplementary Fig. 1b). Recently, a systematic effort, the Genomic Encyclopedia of Bacteria and Archaea (GEBA) Project⁵, has been initiated to maximize coverage of the diversity captured in microbial isolates by phylogenetically targeted genome sequencing. However, GEBA does not address candidate phyla that represent a major unexplored portion of microbial diversity, and have been referred to as microbial dark matter (MDM)⁶.

Metagenomics can obtain genome sequences from uncultivated microorganisms through direct sequencing of DNA from the environment⁷.

In some instances, draft or even complete genomes of candidate phyla have been recovered solely from metagenomic data (Supplementary Table 1). A complementary cultivation-independent approach for obtaining genomes from candidate phyla is single-cell genomics; the amplification and sequencing of DNA from single cells obtained directly from environmental samples⁸. This approach can be used for targeted recovery of genomes and has been applied to members of several candidate phyla (Supplementary Table 1). In particular, natural populations that have a high degree of genomic heterogeneity will be more accessible through single-cell genomics than through metagenomics as co-assembly of multiple strains is avoided. Despite these advances in obtaining genomic representation of MDM, no systematic effort has been made to obtain genomes from uncultivated candidate phyla using single-cell whole genome amplification approaches.

Here, we present GEBA-MDM, the natural extension of the Genomic Encyclopedia into uncultivated diversity by applying single-cell genomics to recover draft genomes from over 200 cells representing more than 20 major uncultivated archaeal and bacterial lineages. Genome-based phylogenetic analysis confirms the validity of rRNA-defined candidate phyla as monophyletic groups and resolves a number of associations among phyla not apparent by single-gene analysis. We discovered several unexpected features, including archaeal sigma factors and stop codon reassignments that challenge established views of the microbial world. Furthermore, we show that single-cell genome references substantially improve the phylogenetic anchoring of about 340 million previously incorrectly or under-classified metagenomic reads.

¹DOE Joint Genome Institute, Walnut Creek, California 94598, USA; ²Center for Biotechnology, Bielefeld University, 33615 Bielefeld, Germany; ³Department of Evolution and Ecology, University of California Davis, California 95616, USA; ⁴Streef Institute, University of Technology Sydney, Ultimo NSW 2057, Australia; ⁵Sigelow Laboratory for Ocean Sciences, East Boothbay, Maine 04544-0380, USA; ⁶Department of Microbiology and Immunology and Graduate Program in Bioinformatics, University of British Columbia, Vancouver, British Columbia, V6T 1Z3 Canada; ⁷School of Life Sciences, University of Nevada, Las Vegas, Nevada 89154-4004, USA; ⁸Department of Environmental and Natural Resources Management, University of Patras, Agrinio, T.K. 30100, Greece; ⁹Biology Department, Woods Hole Oceanographic Institution, Woods Hole, Massachusetts 02543, USA; ¹⁰Department of Civil and Environmental Engineering, University of Illinois at Urbana-Champaign, Urbana, Illinois 61820, USA; ¹¹Australian Centre for Ecogenomics, School of Chemistry and Molecular Biosciences and Institute for Molecular Bioscience, The University of Queensland, St Lucia QLD 4072, Australia;



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Single cell genomics: an individual look at microbes

Ramunas Stepanauskas

Bigelow Laboratory for Ocean Sciences, 60 Bigelow Drive, P.O. Box 380, East Boothbay, ME 04544, United States

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Single cell genomics (SCG) uncovers hereditary information at the most basic level of biological organization. It is emerging as a powerful complement to cultivation-based and microbial community-focused research approaches. SCG has been instrumental in identifying metabolic features, evolutionary histories and inter-organismal interactions of the uncultured microbial groups that dominate many environments and biogeochemical cycles. The SCG approach also holds great promise in microbial microevolution studies and industrial bioprospecting. Methods for SCG consist of a series of integrated processes, beginning with the collection and preservation of environmental samples, followed by physical separation, lysis and whole genome amplification of individual cells, and culminating in genomic sequencing and the inference of encoded biological features.

Highlights

► Single cell genomics (SCG) analyzes DNA at the most fundamental level of biology. ► SCG uncovers metabolic potential of the uncultured microorganisms. ► *In situ* studies of microbial predation, infections and symbioses are enabled. ► SCG will improve our understanding of prokaryote diversity and diversification. ► Genomic bioprospecting of the microbial uncultured majority is enabled by SCG.



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2013

 [Boldly illuminating biology's 'dark matter'. e! Science News, July 2013](#)

 [Bigelow Laboratory Single Cell Genomics — Working to Illuminate Biology's "Dark Matter". Bigelow Laboratory for Ocean Sciences, July 2013](#)

 [Bigelow Scientists Explore Deep Biosphere Under North Pacific Ocean. Bigelow Laboratory for Ocean Sciences, July 2013](#)

 [Jacobson R. New Organisms Shine a Light on 'Microbial Dark Matter'. PBS, July 2013](#)

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 [Hayden EC \(2013\) Single-Cell Sequencing Reveals Genomes of More Than 200 Unusual Microbes. Scientific American, July 2013](#)



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SERVICE DESCRIPTION

Overview

Single cell genomics consists of a series of integrated processes, starting with appropriate collection and preservation of environmental samples, followed by physical separation, lysis, and whole genome amplification of individual cells, then proceeding to either targeted loci or whole genome sequencing and sequence interpretation. SCGC offers a comprehensive suite of single cell genomics services, from single cell separation through genome sequencing and bioinformatics. SCGC also provides advice on environmental sample collection and storage protocols (please see shipping instructions) and post-sequencing analyses (customized services). Most general methods that are currently employed by SCGC have been described previously (e.g. Swan et al. 2011). However, please keep in mind that continued method improvement is a significant component of SCGC activities, and many details of our protocols keep evolving.

A detailed description of SCGC services is available for [download as a PDF file](#).

Click on individual services in the table below for descriptions. References are available for [download](#).

Service	Cat. #	Unit	Bigelow	Other Nonprofit	Corporate
Single Amplified Genome (SAG) Generation	S-101	384-well plate	\$2,700	\$3,200	\$5,400
Bacteria SAG identification	S-102	384-well plate	\$2,200	\$2,600	\$4,400
Archaea SAG identification	S-103	384-well plate	\$2,200	\$2,600	\$4,400
Eukarya SAG identification	S-104	384-well plate	\$2,200	\$2,600	\$4,400
SAG re-arraying	S-105	96-well plate	\$400	\$450	\$800
Prokaryote SAG Whole Genome Sequencing	S-014	1 SAG	\$3,000	\$3,600	\$6,000
SAG re-MDA	S-007	1 SAG	\$400	\$450	\$800
Consultation	S-011	1 hour	free	\$260	\$450
Customized Services	S-100		Req. a quote	Req. a quote	Req. a quote

Human gut flora (bacteria)

- 100 trillion viable bacteria
- 10x total # cells in the body
- Comprise about 2lbs in mass
- Role of many bacteria is not well understood
- Single cell genomics may offer a new and powerful approach





IUCN Information Papers

for the

Intersessional Workshop on Marine Genetic Resources 2-3 May 2013

United Nations General Assembly Ad Hoc Open-ended
Informal Working Group to study issues relating to the
conservation and sustainable use of marine biological
diversity beyond areas of national jurisdiction

Prepared by
IUCN Environmental Law Centre

IUCN Environmental Law Centre
Godesberger Allee 108-112, 53175 Bonn, Germany
Tel: +49 – 228 – 2692 – 231, Fax: +49 – 228 – 2692 – 250
Email: ELCsecretariat@iucn.org, www.iucn.org/law



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